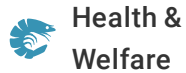




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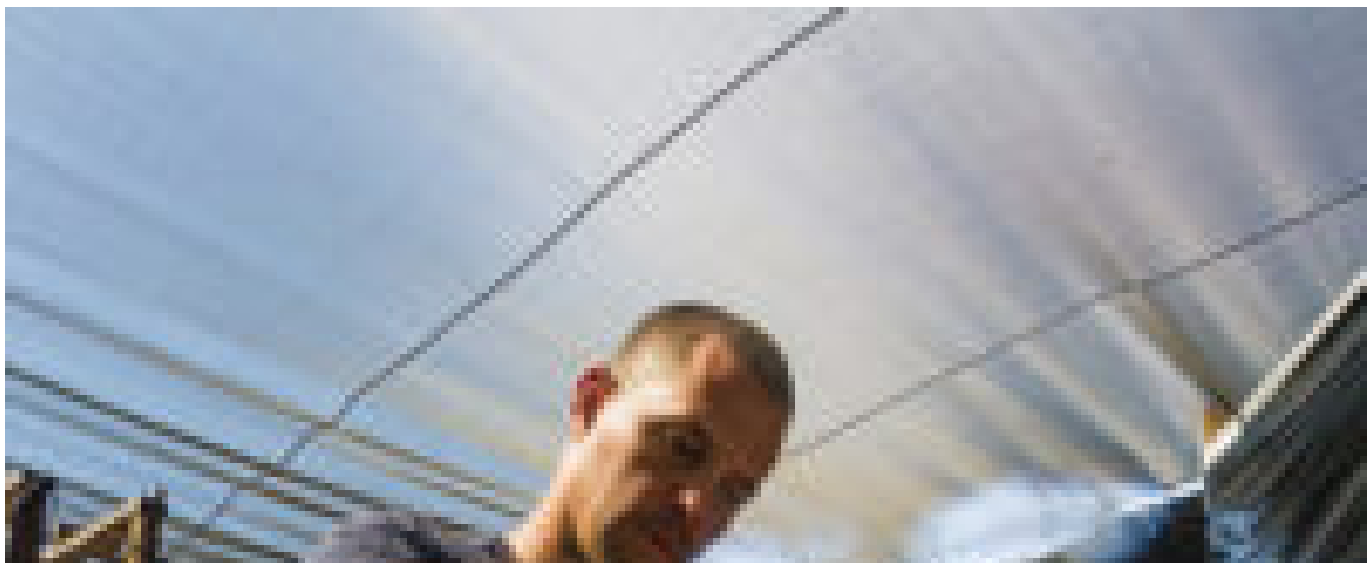
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# Efficacy of saturated lipids in juvenile California yellowtail feeds

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## Understanding, combining LC-PUFA sparing for fish oil replacement





This study compared production performance and tissue fatty acid composition of California yellowtail fed diets with various lipids. Photo by Hubbs-Sea World Research Institute (HSWRI).

The California yellowtail (*Seriola dorsalis*), like many other marine carnivorous fish, has dietary requirements for certain long-chain polyunsaturated fatty acids (LC-PUFAs). More precisely, its feeds must include adequate amounts of arachidonic acid (ARA) and docosahexaenoic acid (DHA). And also including eicosapentaenoic acid (EPA) may also be important for optimizing its growth performance.

Consequently, directly replacing LC-PUFA-rich fish oil with other, less costly alternative lipids typically devoid of these nutrients has proven difficult for *Seriola* spp. Similar information has also been reported for other species with similar essential fatty acid requirements. Adjusting alternative lipid-based feeds with other sources of DHA, EPA and ARA can reestablish performance, but these specialty supplements are costly and reduce the savings resulting through fish oil sparing.

It may be possible to meet the fatty acid requirements of fish like California yellowtail with reduced amounts of costly individual LC-PUFA concentrates if they are fed alternative lipids with little-to-no C18 PUFA content, but results are mixed regarding the relative strength of LC-PUFA sparing associated with SFAs vs. MUFAs. In most, but not all cases, SFA-rich lipids induce a stronger LC-PUFA sparing effect than MUFA-rich lipids, but there has been little work done to assess how much SFA content is enough to induce the sparing effect, whether saturates and unsaturates interact synergistically or antagonistically when both are present in the diet, etc.

In this study, summarized here from the original publication

(<https://doi.org/10.1016/j.aquaculture.2018.04.040>

(<https://doi.org/10.1016/j.aquaculture.2018.04.040>)), we compared production performance and tissue fatty acid composition of California yellowtail fed diets with fish oil or soybean oil (rich in C18 PUFAs), partially hydrogenated soybean oil (rich in MUFAs), fully hydrogenated soybean oil (rich in SFAs), or blends of these soybean oils supplemented to meet requirements for LC-PUFAs previously determined.



View of the study experimental setup. Photo by Hubbs-Sea World Research Institute (HSWRI).

## Study setup

Juvenile California yellowtail (initial individual weight= $6.2 \pm 0.0$  grams, mean  $\pm$  SEM) were randomly stocked in a semi-closed recirculating aquaculture system equipped with supplemental aeration, mechanical and biological filters at the Hubbs-Sea World Research Institute (HSWRI) facility. Twenty-four, 300-L tanks were stocked with 15 fish/tank, and dietary treatments were randomly assigned to quadruplicate tanks.

The experiment lasted for 10 weeks. Fish were fed continuously with belt feeders, at a ration of approximately 7 percent of body weight/day (equivalent to apparent satiation) equally divided into six feeding events/day. Tanks were monitored regularly for uneaten feed and none was observed during the daily siphoning events to remove solids. All animal husbandry and sampling protocols, including those described below, were in keeping with the HSWRI standards and guidelines for humane animal care and use in research.

Dietary treatments were formulated to contain 473.7 grams per kg of menhaden fish meal and 240 grams per kg of soy protein concentrate. Experimental diets varied only in lipid source and fatty acid composition (e.g. fatty acid groupings such as SFAs, MUFAs and C18 PUFAs) The control diet ("FISH") was based on previous tested formulations and contained menhaden fish oil as the only supplemental lipid source.

The experimental dietary treatments contained various types of soy-derived lipids as needed to achieve distinct fatty acid profiles favoring different fatty acid groupings, including fully hydrogenated SFA-rich soybean oil ("SFA SOY" diet); partially hydrogenated MUFA-rich soybean oil ("MUFA SOY" diet); standard C18 PUFA-rich soybean oil ("C18 PUFA SOY" diet); a blend of fully and partially hydrogenated soybean oils providing similar levels of SFAs and MUFAs, but low C18 PUFA content ("LOW PUFA BLEND" diet); and another blend containing all soybean oil types providing an approximately equal balance of SFAs, MUFAs, and C18 PUFAs ("BALANCED SOY BLEND").

To ensure the LC-PUFA requirements of juvenile California yellowtail were met, the soybean oil-based experimental diets were supplemented with LC-PUFA concentrates rich in EPA, DHA and ARA to achieve the necessary levels of these fatty acids; because of the EPA, DHA, and ARA content of menhaden fish oil, the FISH diet did not require supplementation.

The experimental diets used in the study were prepared at the Center for Fisheries, Aquaculture and Aquatic Sciences (CFAAS; Carbondale, Ill., USA) according to standard in-house feed manufacturing procedures. After collecting sub-samples of each diet for compositional analysis diets were sent to the Hubbs-Sea World Research Institute (HSWRI; Carlsbad, Calif., USA) facilities for the feeding trial where they were kept frozen prior to use (minus-20 degrees-C).

For detailed descriptions of the procedures for experimental design and feeding trial; experimental feed preparation and analyses; production performance, sample collection and analyses; tissue fatty acid analyses and coefficient of distance (Djh) values; statistical analyses; and references, please contact the first author.



All dietary lipid sources, including menhaden fish oil and the various types of soy-derived lipids with distinct fatty acid profiles, were readily accepted by the juvenile yellowtail and supported normal growth and production performance. Photo by Hubbs-Sea World Research Institute (HSWRI).

## Results and discussion

Overall results show that fish survival during the trial was 100 percent, and that production performance was not affected by dietary lipid source or fatty acid composition (Table 1). Fish growth was relatively fast, with weight gain averaging  $1442 \pm 20$  percent (grand mean  $\pm$  SEM) and SGR averaging  $3.91 \pm 0.02$  percent body weight/day across all treatment groups. Feed intake (6.59 to 6.82 percent body weight/day) and FCR (1.25 to 1.33) were also unaffected by dietary treatment. And the organosomatic indices were not significantly different, and differed little among the treatment groups.

### Rombenso, CYT, Table 1

Production variable	FISH	SFA SOY	MUFA SOY	C18 PUFA SOY	LOW PUFA BLEND	BALANCED SOY BLEND	PSE	P value
Initial weight (g)	6.2	6.2	6.22	6.3	6.2	6.3	0.1	0.566
Final weight (g)	96.2	95.5	97.5	91.5	99.2	98.4	3.2	0.258
Weight gain (%)	1454	1431	1470	1351	1488	1457	50	0.156
FCR	1.3	1.26	1.26	1.33	1.25	1.29	0.03	0.356
SGR (% body weight/day)	3.92	3.90	3.93	3.82	3.95	3.93	0.05	0.127

Feed intake (% body weight/day)	6.82	6.59	6.69	6.71	6.68	6.80	0.11	0.316
HIS	10	0.9	1.0	1.0	0.9	1.0	0.1	0.925
VSI	7.2	7.1	7.0	7.0	7.1	7.0	0.4	0.982

Table 1. Production performance by dietary treatment. Values represent least-square means; pooled SE (PSE) values and P values resulting from one-way ANOVA tests are also provided; means with common letter labels are not significantly different ( $P > 0.05$ ).

All dietary lipid sources, including menhaden fish oil and the various types of soy-derived lipids with distinct fatty acid profiles, were readily accepted by the juvenile yellowtail and supported normal growth and production performance. This is consistent with the fish oil sparing literature that indicates that nearly any lipid source could be suitable for use in aquafeeds if the essential fatty acid requirements of the targeted species are fulfilled.

In our previous work, we demonstrated the importance of ARA, DHA, and (to a lesser extent) EPA in satisfying the essential fatty acid requirements of California yellowtail, so these were provided directly in the form of lipid supplements to the soy lipid-based feeds. Although the requirements for growth and survival were met, it was expected that changes in dietary lipid source and composition would result in changes in tissue fatty acid profiles. In fact, tissue fatty acid profiles did generally mimic dietary fatty acid composition as reported in the literature.

It is becoming more evident that fatty acid composition of dietary alternative lipid is important. Specifically, the abundance of SFAs vs. unsaturated fatty acids influences the magnitude of tissue fatty acid distortion and it is possible to manipulate tissue fatty acid composition, including LC-PUFA levels, using dietary lipids with certain fatty acid profiles.

Lipids rich in C18 PUFA seems to negatively interfere with LC-PUFA availability and deposition in tissues, whereas those rich in SFAs and/or MUFAs have been described as advantageous in the context of preserving tissue fatty acid composition and enhancing availability of LCPUFA. Although few studies reported no such benefits of SFA- and MUFA-rich lipids, this trend appears to be valid for a number of freshwater and marine taxa.

Additionally, in some circumstances SFA-rich diets appear to substantially reduce dietary requirements for LC-PUFAs and dietary supplementation levels to enhance tissue deposition of these nutrients. This mechanism, in which LC-PUFA availability and deposition are correlated with dietary SFA and MUFA, has been described as the omega-3 or LC-PUFA “sparing effect.”

Fatty acids with shorter chain lengths and lower degrees of unsaturation appear to be preferentially targeted for catabolic shunting. The most abundant fatty acids in SFA- and MUFA-rich lipids used in this study were 18:0 and 18:1n-9, respectively. Presumably a catabolic preference for 18:0 over 18:1n-9 contributed to the underrepresentation of 18:0 in the tissues for (or “resulting in”) at least part of the sparing effect advantage of the SFA-rich feeds over the

MUFA-rich feeds. Regardless of the mechanism(s) responsible, SFAs appear to provide a distinct benefit beyond that associated with MUFAs in terms of conserving tissue fatty acid profiles and LC-PUFA levels.

So far, most studies have evaluated the benefits of dietary SFAs and MUFAs separately. Our study clearly demonstrated the benefits of SFA-rich diets over MUFA-rich diets in the context of preserving tissue fatty acid profile and LC-PUFA deposition. The degree of overall tissue fatty acid distortion was softened by dietary SFA, increasing dietary levels of SFAs increased profile similarity to FISH control diet.

The dietary treatments we tested significantly affected fillet total lipid content. Lower lipid levels were observed in the SFA SOY and LOW PUFA BLEND groups, while the FISH and C18 PUFA SOY groups had the highest lipid levels in fillet. This observation – fish oil-based diets yielding higher total lipid content in fillet compared to alternative and complementary lipid sources – has also been reported in other fish species like Atlantic salmon and white sea bass, but not in others like Florida pompano, European sea bass and cobia.

Seemingly, the effect of dietary fatty acid composition on fillet lipid level might be a species-specific response. The dietary fatty acid profile generally influences tissue lipid class composition, in particular neutral lipids, which seem to mimic the dietary fatty acid composition; whereas, polar lipids are more selective exhibiting a preference for certain fatty acids. Although we did not investigate lipid classes, the lipid class composition of fillet was likely modified due to significant changes in the total lipid content.

Results of the study indicate that dietary SFAs and MUFAs interact antagonistically, that MUFAs are inferior to SFAs in terms of LC-PUFA sparing, and that SFAs are the primary drivers in the LC-PUFA “sparing effect” mechanism. Photo by Hubbs-Sea World Research Institute (HSWRI).

## Perspectives

Based on results of our study, there is a stronger relationship between dietary levels of PUFAs (negative) and SFAs (positive) and tissue fatty acid profile composition, including LC-PUFA content, than dietary levels of MUFAs.

We also found that SFA-rich lipids maintained tissue profile and produced equivalent or higher fillet levels of LC-PUFAs compared to the LC-PUFA-rich FISH diet, whereas MUFA- and C18 PUFA-rich lipids contributed, to a lesser and greater magnitude, respectively, towards tissue profile distortion at the expense of LC-PUFAs.

Consequently, we propose that dietary SFAs and MUFAs interact antagonistically, that MUFAs are inferior to SFAs in terms of LC-PUFA sparing, and that SFAs are the primary drivers in the LC-PUFA “sparing effect” mechanism.

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