



ALLIANCE™

(<https://debug.globalseafood.org>).

---



Health &  
Welfare

---

# Study of co-infection of WSSV and *Vibrio parahaemolyticus*

11 November 2019

By Jee Eun Han, DVM, Ph.D. , Ji-Eun Kim, MS , Hayun Jo, MS , Jong-Su Eun, Ph.D. , Chorong Lee, Ph.D. , Ji Hyung Kim, Ph.D. , Kyeong-Jun Lee, Ph.D. and Jae-Won Kim, Ph.D.

## Results show a risk factor for AHPND in Pacific white shrimp



Results of this study demonstrated that shrimp initially exposed to WSSV allow VpAHPND to cause faster and higher mortality than in shrimp infected with VpAHPND alone. Photo by Fernando Huerta.

Acute hepatopancreatic necrosis disease (AHPND), caused by specific strains of the bacterium *Vibrio parahaemolyticus* (Vp<sub>AHPND</sub>), has been reported in shrimp cultured in Southeast Asia and the Americas since 2009. The clinical signs of the disease include atrophied hepatopancreas (HP), an empty gastrointestinal tract, a milky appearance of the stomach and lethargy. This bacterial disease has caused up to 100 percent mortality in populations of farmed Pacific white shrimp (*Penaeus vannamei*) and black tiger shrimp (*P. monodon*), and collective economic losses due to AHPND and other shrimp diseases are estimated at \$23.6 billion globally with a further loss of \$7 billion in feed sales.

In some cases, Vp<sub>AHPND</sub> has been reported in co-infection with the spore-forming microsporidian *Enterocytozoon hepatopenaei* (EHP). Although EHP does not appear to cause mortality, EHP-infected animals would be more vulnerable to AHPND, which may cause substantial mortality in shrimp ponds. And EHP-infected shrimp have an increased susceptibility to Vp<sub>AHPND</sub>, suggesting that shrimp weakened by EHP would succumb to secondary pathogens, such as Vp<sub>AHPND</sub>.

For several years, white spot syndrome virus (WSSV) has spread worldwide throughout much of Asia, the Americas, Europe, Africa, Middle East and Australia. In some cases, WSSV has been detected in healthy animals without clinical symptoms or mortalities, indicating that infection by WSSV does not necessarily result in high mortality, and secondary infection by opportunistic pathogens, such as *Vibrio* spp., would expedite the onset of the disease.

In 2017, *P. vannamei* mortality rates of up to 60 percent were reported in the Philippines. The shrimp exhibited clinical signs of AHPND but were confirmed by PCR to be positive for both Vp<sub>AHPND</sub> and WSSV. From this case, we hypothesized that WSSV infection without clinical symptoms would allow Vp<sub>AHPND</sub> to cause faster and higher mortality of pond cultured shrimp than those infected with Vp<sub>AHPND</sub> alone.

This article – adapted and summarized from **the original** (<https://doi.org/10.1016/j.aquaculture.2019.734333>) – reports on a study designed to support this hypothesis by simulating the co-infection with WSSV and Vp<sub>AHPND</sub> in juvenile shrimp *P. vannamei* under laboratory conditions. As a result, we demonstrated that shrimp initially exposed to WSSV were easily infected by Vp<sub>AHPND</sub>, but hardly recovered from the disease according to histopathology examination. In addition, using qPCR assay and immunohistochemistry [IHC; the most common application of immunostaining, involving the process of selectively identifying antigens (proteins) in cells of a tissue section using the binding of antibodies specifically to antigens in biological tissues] examination, we determined that WSSV infection could be accelerated by the secondary Vp<sub>AHPND</sub> infection. This work was supported by the National Research Foundation of Korea (NRF) grant funded by the Korea government (MSIT) (grant number NRF-2018R1C1B5086350).

## Study setup

Postlarval *P. vannamei* were sourced from a local shrimp farm (Jeju Province, South Korea) and transported to the Institute of Marine Sciences of Jeju National University in South Korea. Shrimp were reared to an average weight of  $0.5 \pm 0.05$  grams in 96-liter maintenance tanks with aerated artificial seawater at 25 to 28 degrees-C and 30 ppt salinity. For the co-infection experiment, juvenile shrimp (average 0.5 grams, N=80) were randomly selected from the maintenance tanks and distributed into experimental tanks. Before the co-infection experiment, representative shrimp were randomly selected and confirmed to be negative for both WSSV and Vp<sub>AHPND</sub> by PCR.

For WSSV stocks, moribund *P. vannamei* with white spots on their cuticles were collected from an anonymous shrimp farm in Chungcheong Province located on the western coast of Korea. For Vp<sub>AHPND</sub> bacterial stocks, the strain 13-028/A3 was previously isolated from the stomach of moribund *P. vannamei* from a farm in Vietnam and displaying whitish hepatopancreases (HP).

For detailed information on the experimental designs for WSSV and Vp<sub>AHPND</sub> co-infection; qPCR assays; IHC examination of the gill tissues; histopathology examination of the HP tissues; and statistical analyses, please refer to the original publication.

## Results and discussion

We studied co-infection with WSSV and Vp<sub>AHPND</sub> in juvenile *P. vannamei* shrimp under laboratory condition, and the mortality rates were compared with single infection groups.

Our results showed that co-infection accelerated shrimp mortality compared with single infection (Fig. 1). We observed 0 percent mortality Group 1 (WSSV only), and cumulative mortality was 50 percent at day 4 post-infection (p.i.) in Group 2 (Vp<sub>AHPND</sub> only). However, the mortality rate increased significantly in Group 3 (WSSV+Vp<sub>AHPND</sub>) compared with the other groups ( $p < .05$ ). In Group 3, the shrimp began to show mortality at day 2 p.i., and cumulative mortality reached 75 percent at day 3 p.i. During the 10-day experimental period, no mortality was observed in the negative control (Group 4, not challenged).

Our results also showed that co-infection increased the level of WSSV infection, which was confirmed via qPCR and IHC, and the levels of WSSV infection were markedly different between Group 1 (WSSV only) and Group 3 (WSSV + Vp<sub>AHPND</sub>) on the termination day (Day 10). Based on qPCR analysis, WSSV was detected in shrimp in Group 3 (WSSV+Vp<sub>AHPND</sub>) but WSSV was negative in shrimp exposed to WSSV only (Group 1). Also, by IHC examination, brown coloration against pink cytoplasm and purple nuclei, indicative WSSV-positive reactions were observed in the shrimp of Group 3 (WSSV+Vp<sub>AHPND</sub>) (Fig. 2D) similar to the WSSV-positive sample previously prepared in the preliminary study (Fig. 2B). However, these WSSV-positive reactions were not seen in the shrimp exposed to WSSV alone (Group 1;

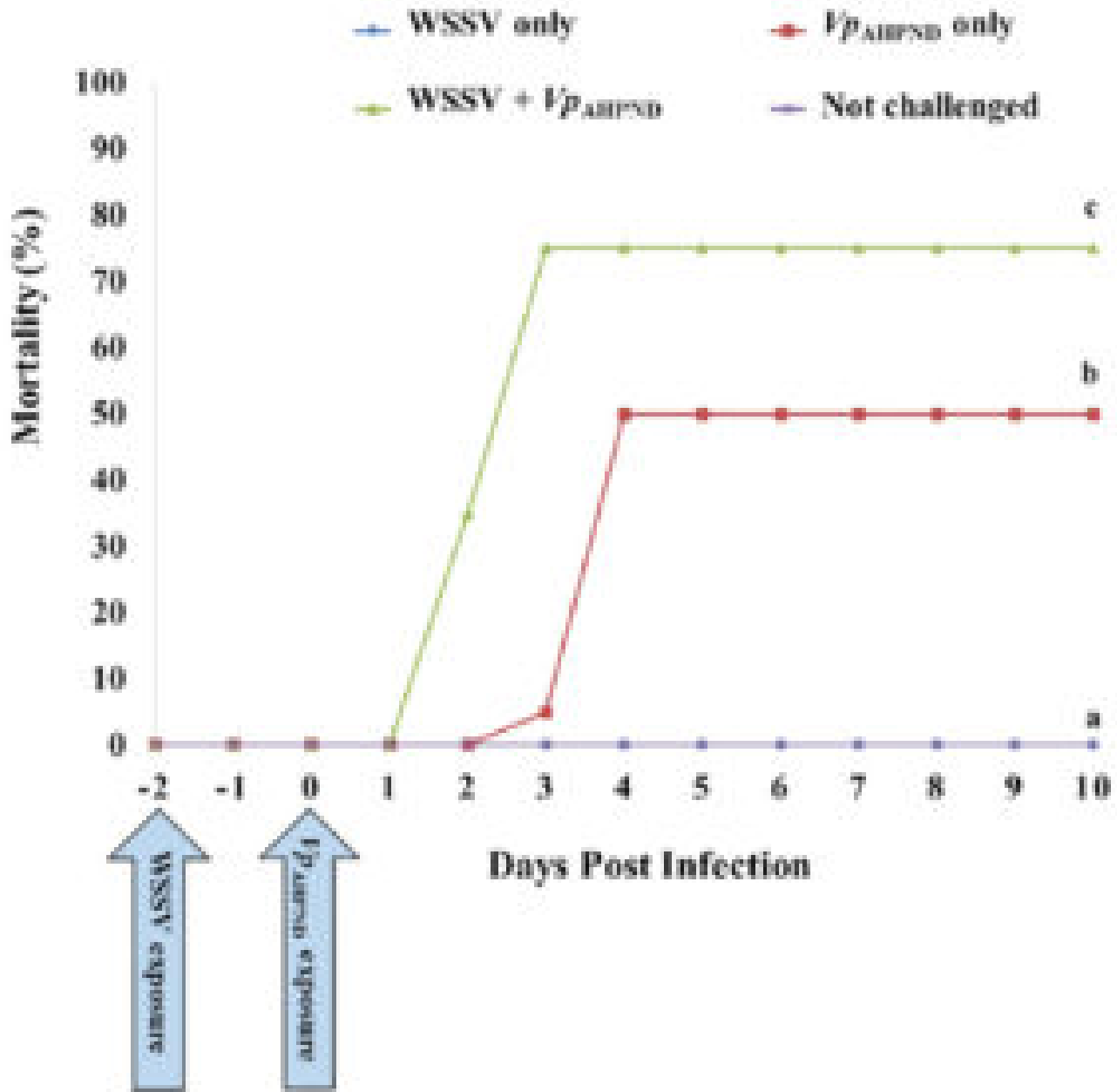


Fig. 1: Cumulative mortality of *P. vannamei* after challenge with WSSV and *Vp*<sub>AHPND</sub> (Group 1: WSSV only; Group 2: *Vp*<sub>AHPND</sub> only; Group 3: WSSV + *Vp*<sub>AHPND</sub>; Group 4: not challenged). The mortality rate was significantly increased in the co-infection group (Group 3) compared with the single infection groups (Groups 1 and 2). Different letters indicate significant differences in survival at the termination day (day 10) ( $P < .05$ ).

Fig. 2A) and the negative control (Group 4, not challenged; Fig. 2C).

Co-infection decreased the recovery of *Vp*<sub>AHPND</sub> infection. All experimental shrimp showed typical clinical symptoms of AHPND, including pale HP and empty gut, within three to four days after infection in Group 2 (*Vp*<sub>AHPND</sub> only) and Group 3 (WSSV+*Vp*<sub>AHPND</sub>), and *Vp*<sub>AHPND</sub> infection was confirmed by

qPCR and histopathology examination. From the qPCR analysis,  $Vp_{AHPND}$  was detected in dead shrimp collected from both Group 2 ( $Vp_{AHPND}$  only) and Group 3 (WSSV+ $Vp_{AHPND}$ ) on Day 3 and 4.

By histopathology examination, representative shrimp from both Group 2 ( $Vp_{AHPND}$  only) and Group 3 (WSSV +  $Vp_{AHPND}$ ) showed a typical terminal phase, characterized by massive bacterial infection and sloughing of hepatopancreatic tubule epithelial cells at the level of G3-4 (grade 4 is the most severe) on Day 3.

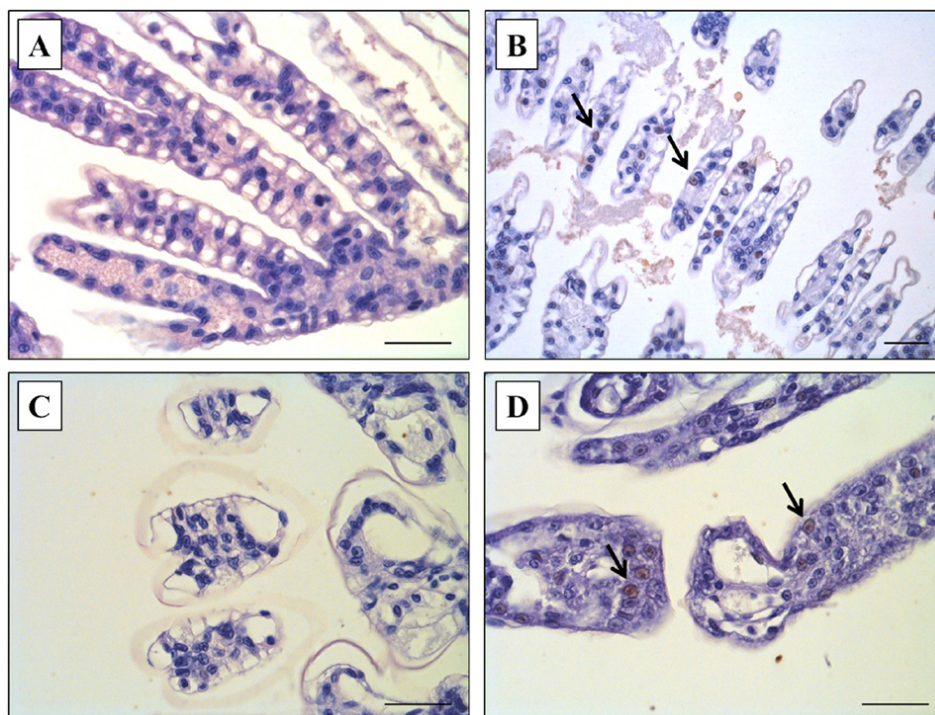


Fig. 2: Immunohistochemistry (IHC) examination from shrimp infected with white spot syndrome virus (WSSV). (A) Negative control (Group 4): no WSSV-infected cells in gills of *P. vannamei*. (B) Positive control: WSSV-positive cells in gills of *P. vannamei* infected with WSSV tissue by the feeding method. (C) WSSV control (Group 1): no WSSV-infected cells in gills of *P. vannamei* infected with WSSV by the immersion method. (D) WSSV and  $Vp_{AHPND}$  co-infection (Group 3): WSSV-positive cells in gills of *P. vannamei* pre-infected with the WSSV and post-challenged with the secondary pathogen,  $Vp_{AHPND}$ . Hematoxylin counterstain. Scale bar: 40  $\mu$ m.

However, on the termination day (day 10), a remarkable difference in the histopathology of the HP of surviving shrimp was seen between group 2 ( $Vp_{AHPND}$  only) and Group 3 (WSSV+ $Vp_{AHPND}$ ). Histopathology signs of the group 2 ( $Vp_{AHPND}$  only) survivors had an HP of normal appearance, and normal functions of B and R cells (Fig. 3B), the same as the negative control (Group 4, Fig. 3A). To the contrary, Group 3 (WSSV+ $Vp_{AHPND}$ ) survivors showed typical AHPND histopathology signs in the HP, with massive bacterial infection and sloughing of hepatopancreatic tubule epithelial cells of G4 severity (Fig. 3C).

Fig. 3: Histopathology examination using Davidson's AFA-fixed tissue sections from shrimp infected with VpAHPND, on the termination day (Day 10). (A) Negative control (Group 4): normal appearance of the hepatopancreas. (B) AHPND control (Group 2): normal appearance of the hepatopancreas with normal function of B and R cells. (C) WSSV and VpAHPND co-infection (Group 3): the typical acute phase with progressive necrosis and massive sloughing of hepatopancreatic tubule epithelial cells. Mayer-Bennett's hematoxylin/eosin-phloxine (H&E) staining. Scale bar: 180  $\mu$ m.

Infections with multiple pathogens (co-infection) are common in practical shrimp cultures. Several cases of shrimp vibriosis with WSSV infection have been reported with various penaeid species in different countries. There has been no prior detailed analysis on the roles of these pathogens in shrimp co-infected with WSSV and *Vibrio*. In shrimp farms, the WSSV infection without symptoms or mortality probably causes shrimp weakening and might trigger the VpAHPND infection.

We also observed declines in antioxidant and antiperoxidative enzymes in shrimp infected with VpAHPND, similar to WSSV in our ongoing study, and we suspect that VpAHPND infection may also have compromised the animals, increasing their susceptibility to other pathogens in ponds.

Intramuscular (IM) routes have been widely used for experimental infection of shrimp with WSSV. However, IM injection has not been considered as an infection route of VpAHPND. In addition, per os feeding can produce both WSSV and VpAHPND infection, but this is not a practical method for the dual-infection experiments because experimental shrimp were supposed to be infected twice within short intervals. Therefore, for the current bioassay, we selected the immersion (waterborne) method to co-infect shrimp with WSSV and VpAHPND. The synergistic effects influenced by the same infection routes have already been confirmed between WSSV and *Vibrio* spp.

## Perspectives

To the best of our knowledge, our study is the first to examine the co-infection of WSSV and VpAHPND under laboratory conditions. In ponds, shrimp that were infected with WSSV (or other pathogens) did not always show clinical signs or mortalities. In those cases, even WSSV-positive shrimp without any clinical symptoms may cause higher and faster mortality than their disease-free counterparts, and the shrimp would hardly recover from the disease following a secondary infection, such as VpAHPND.

Also, the secondary infection would provoke WSSV infection or virus replication, as confirmed in the present study. Therefore, shrimp farmers should pay attention to the management of multiple infections (WSSV or other pathogens) in their ponds to prevent further losses in shrimp production.

*References available in the original publication.*

## Authors

---



**JEE EUN HAN, DVM, PH.D.**

Laboratory of Aquatic Biomedicine  
College of Veterinary Medicine  
Kyungpook National University  
Daegu 41566, Korea

[hanje1223@gmail.com](mailto:hanje1223@gmail.com) (<mailto:hanje1223@gmail.com>).



**JI-EUN KIM, MS**

Institute of Integrated Technology  
CJ CheilJedang, CJ Blossom Park  
Suwon 16495, Republic of Korea



**HAYUN JO, MS**

Institute of Integrated Technology  
CJ CheilJedang, CJ Blossom Park  
Suwon 16495, Republic of Korea



**JONG-SU EUN, PH.D.**

Institute of Integrated Technology  
CJ CheilJedang, CJ Blossom Park  
Suwon 16495, Republic of Korea



**CHORONG LEE, PH.D.**

Department of Marine Life Sciences  
Jeju National University  
Jeju 63243, Republic of Korea



**JI HYUNG KIM, PH.D.**

Infectious Disease Research Center  
Korea Research Institute of Bioscience and Biotechnology  
Daejeon 34141, Republic of Korea



**KYEONG-JUN LEE, PH.D.**

Department of Marine Life Sciences  
Jeju National University  
Jeju 63243, Republic of Korea



**JAE-WON KIM, PH.D.**

Institute of Integrated Technology  
CJ CheilJedang, CJ Blossom Park  
Suwon 16495, Republic of Korea



